



A Comparative Analysis of Recombinant AAV9 Product Generated from Insect and Mammalian Bioproduction Processes

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Disclosures

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Background

- Mammalian (HEK-AAV) and insect cell-based (Sf9-AAV) manufacturing systems are the two predominant AAV manufacturing platforms
- Neurogene has established both manufacturing platforms and have cleared INDs with each process

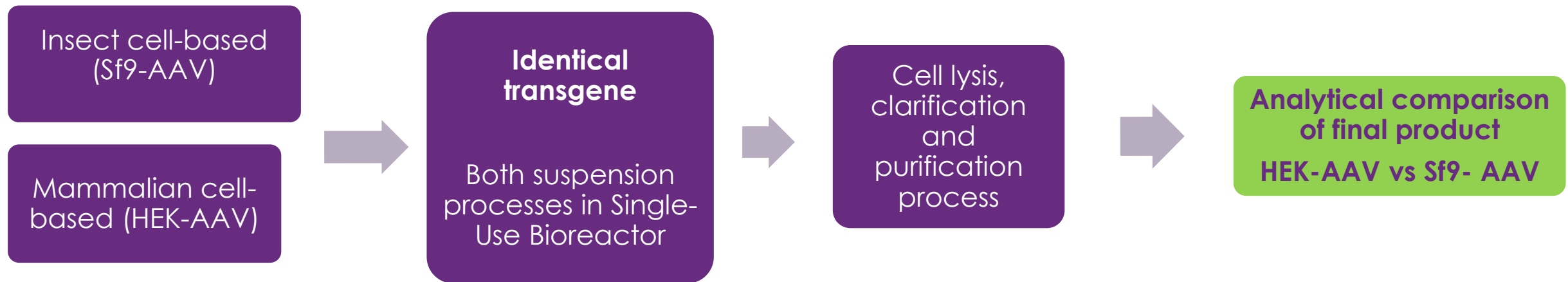
| | Sf9-AAV | HEK-AAV |
|------------|--|---|
| Advantages | <ul style="list-style-type: none"> • Higher productivity and lower COGS • Robust scale-up • Better safety profile (absence of proto-oncogene in production cells, less rcAAV) • Little or no expression of transgenes in insect cells | <ul style="list-style-type: none"> • Flexibility to switch from one serotype and/or transgene to another • Speed and established protocols to generate material |
| Challenges | <ul style="list-style-type: none"> • Requires master and working banks of both recombinant baculovirus clones (upfront time and resource utilization) • Might require viral clearance demonstration in early phases (even with Rhabdo-free cell line) | <ul style="list-style-type: none"> • Lower productivity and higher COGS • Scale-up challenges: Requires carefully controlled mixing at transfection step • Some transgene expression may affect performance of the cell culture system |



AAV = adeno-associated virus; COGS = cost of goods sold; rcAAV = replication competent AAV; Sf9 = *Spodoptera frugiperda*; HEK = human embryonic kidney

Study Objective- Analytical Comparison of Mammalian and Insect Cell-based Manufacturing Systems

Two optimized, scalable platforms were utilized to generate AAV9 containing same **transgene of medical importance (GMI)**

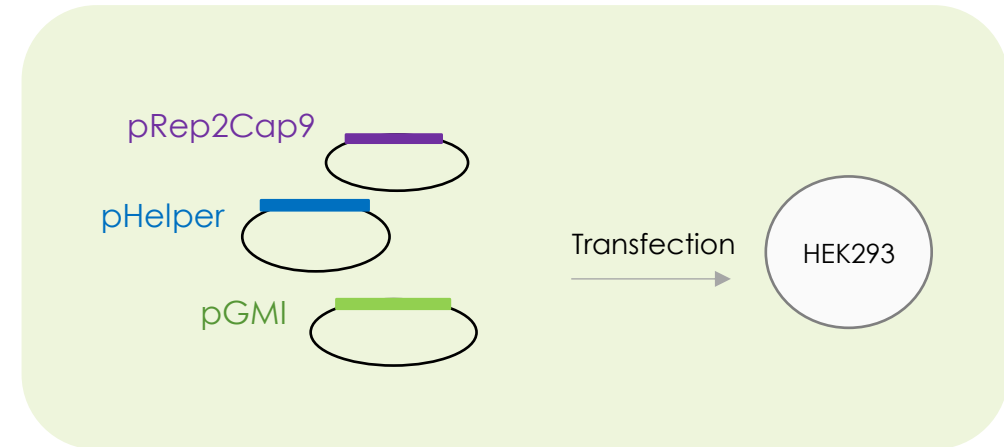
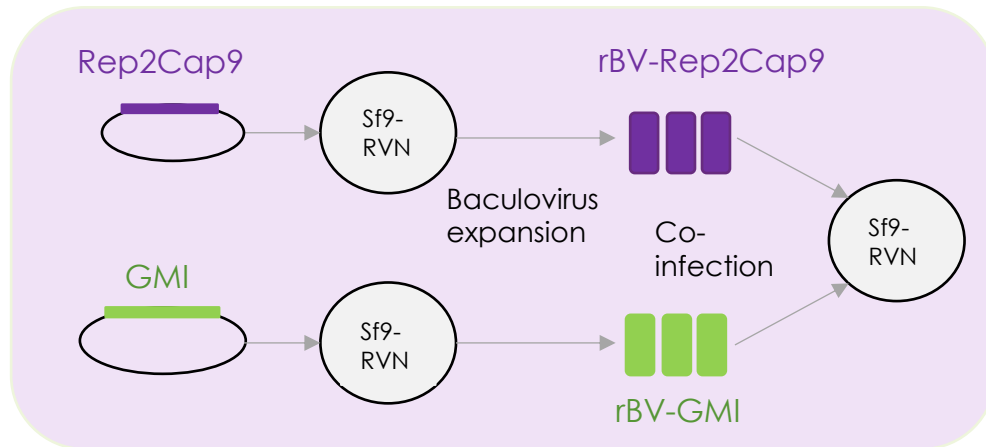


Process Overview of AAV9 Production Systems

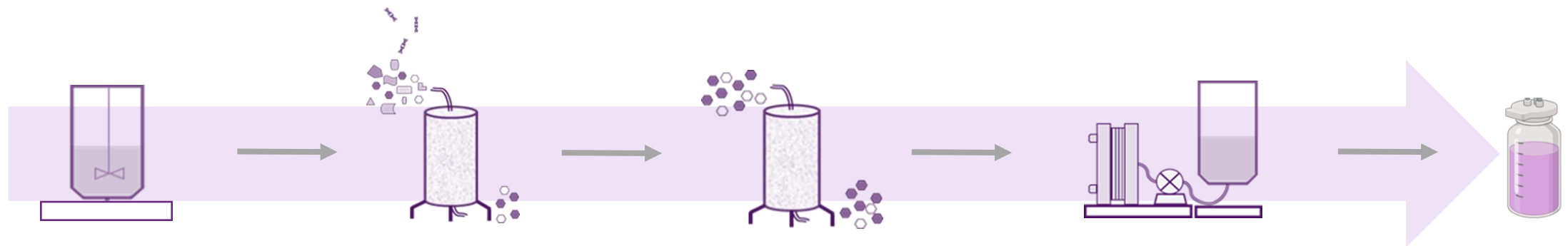
Sf9-AAV

HEK-AAV

Upstream



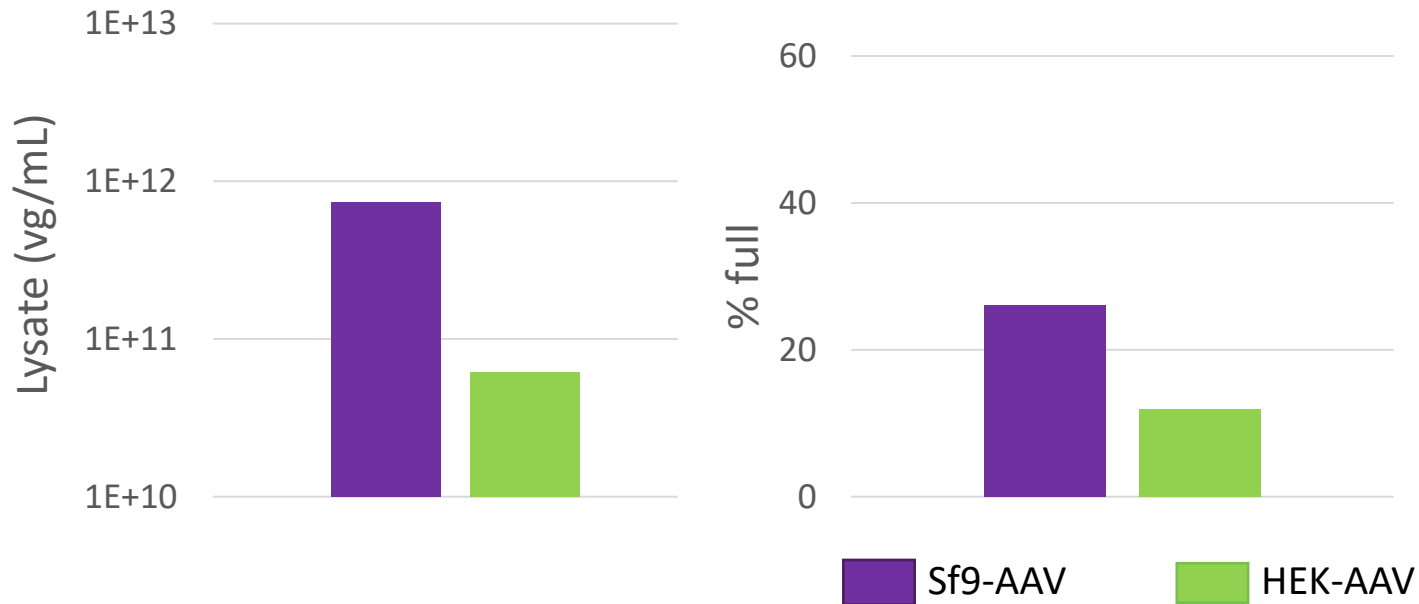
Downstream



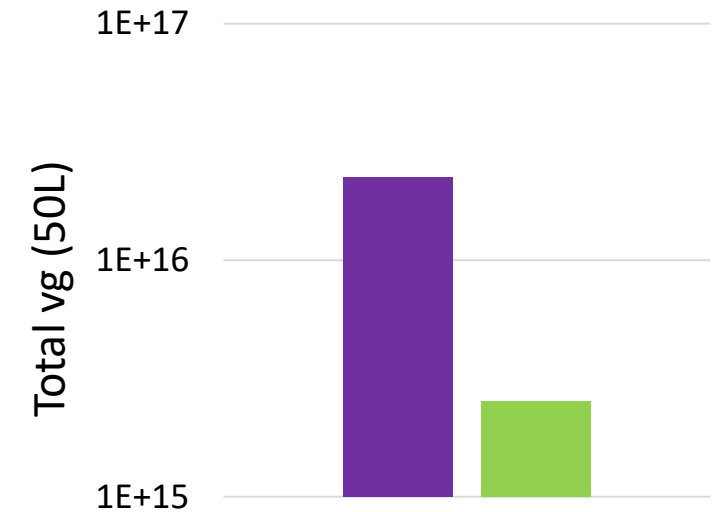
Same cell lysis, clarification, and purification processes

The Insect Cell-based System Yields Higher Productivity and Percent Full at Harvest

Productivity and % full at harvest



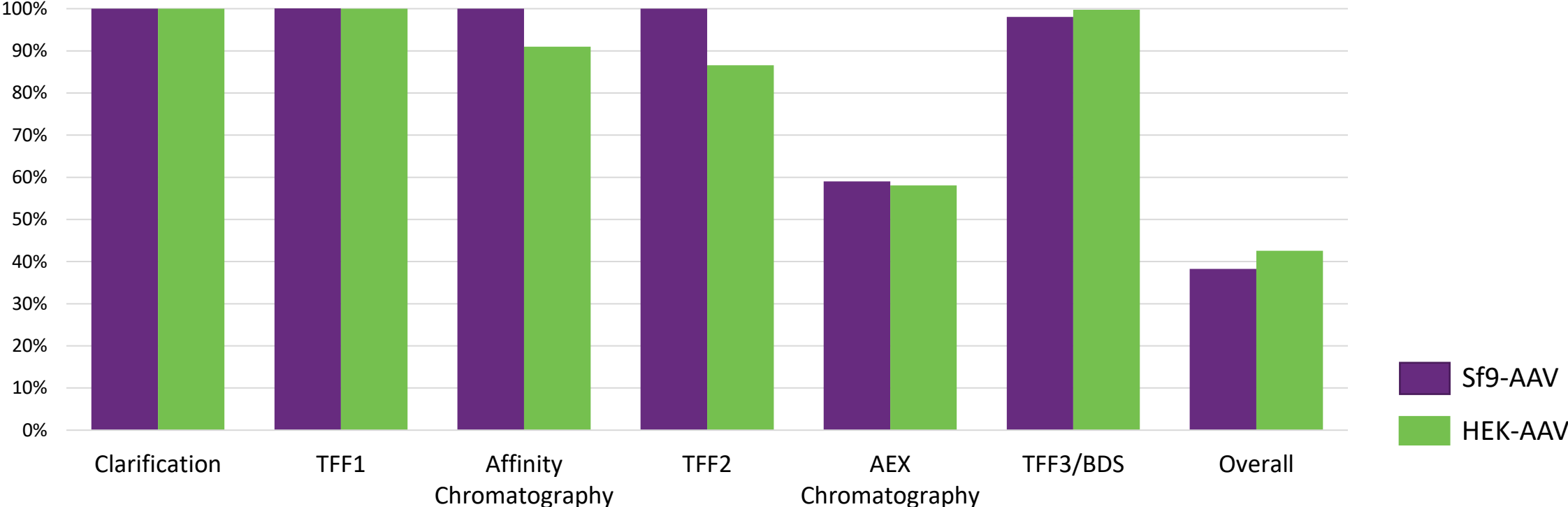
Volumetric yield



Total yield from the same scale runs is ~10-fold higher using the Sf9-AAV system

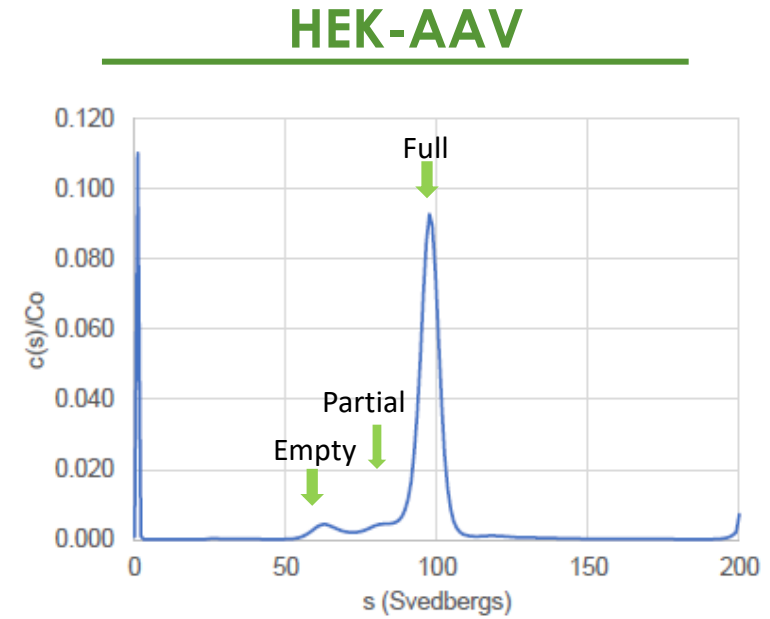
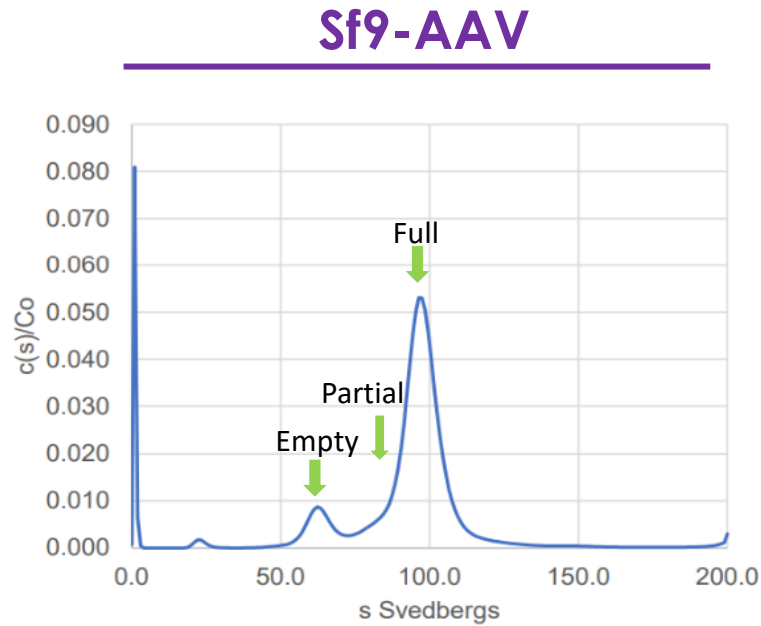
Recoveries From Each Unit Operation are Similar Between the Two Processes

Sf9-AAV and HEK-AAV Step recoveries by ddPCR



TFF = tangential flow filtration; AEX = anion exchange; BDS = bulk drug substance; ddPCR = droplet digital polymerase chain reaction

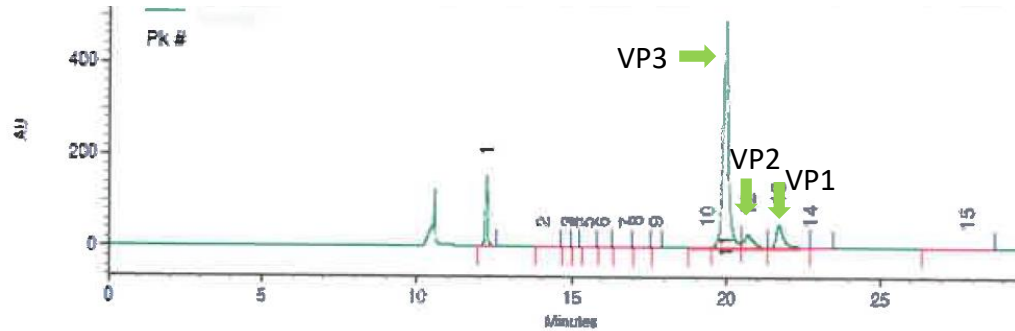
Both Processes Resulted in Similar AAV Particle Content by AUC



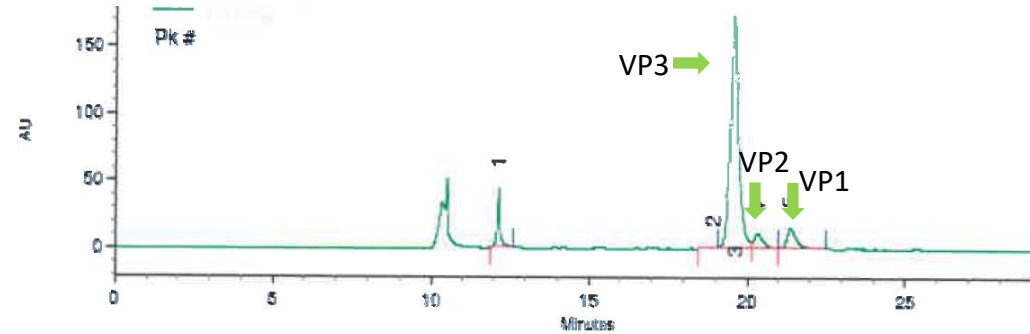
| Particle Content (%) | Sf9-AAV | HEK-AAV |
|----------------------|---------|---------|
| Empty | 10 | 6 |
| Partial | 8 | 7 |
| Full | 82 | 87 |

Similar Capsid Composition (Viral Protein Ratio) Observed in Both Products by CE-SDS

Sf9-AAV

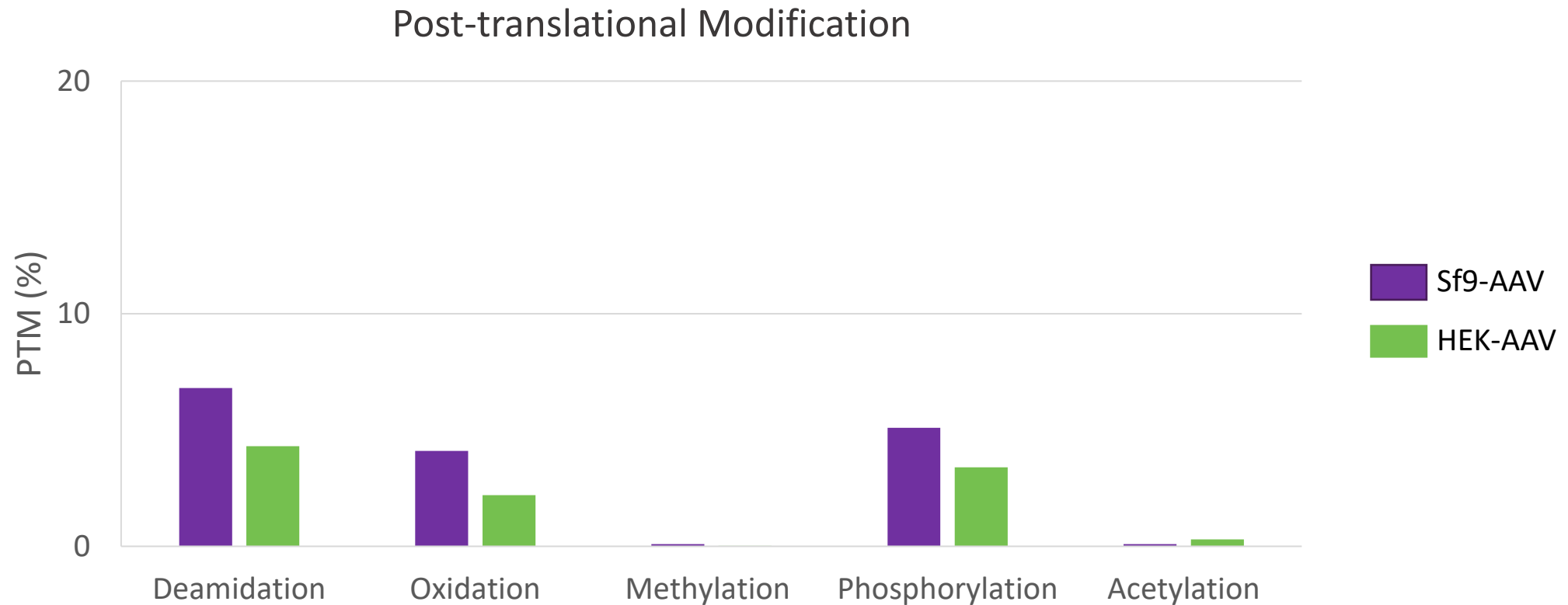


HEK-AAV



| Species | VP1 | VP2 | VP3 | AAV Purity (%) |
|---------|-----|-----|-----|----------------|
| Sf9-AAV | 1.5 | 1.0 | 10 | 90 |
| HEK-AAV | 1.1 | 0.8 | 10 | 93 |

Overall Low Levels of PTM on the Capsid Surface, and the Difference between Products is within Assay Variability



MiSeq Data Analysis Showed Similar Genome Integrity for Both Processes

| Regions | Sf9-AAV (reads aligning to map %) | HEK-AAV (reads aligning to map %) |
|------------------------------|--------------------------------------|--------------------------------------|
| NGN Construct (GMI) | 86 | 91 |
| Starting Plasmid Backbone | 0.02 | 1.30 |
| Baculo RepCap/Plasmid RepCap | 0.18 | 0.48 |
| Shuttle Vector | 0.010 | N/A |
| Helper Plasmid | N/A | 0.21 |
| Host Cell DNA | 1.10 | 0.57 |

Residual (Impurity) Analysis and Safety Testing Showed Comparable Profiles

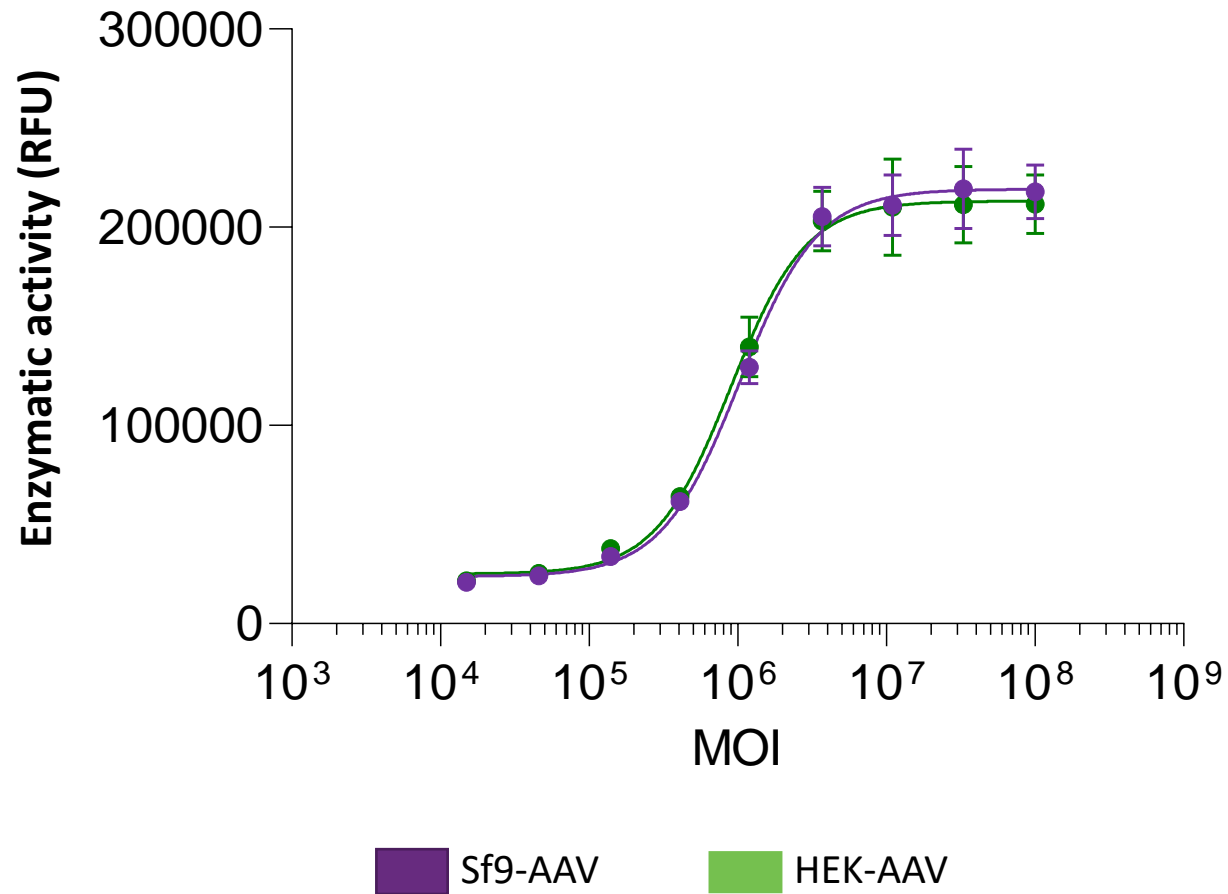
| Assay | Sf9-AAV | HEK-AAV |
|---|------------------------------|------------------------------|
| Endotoxin (EU/mL) | < 0.05 | < 0.05 |
| SEC (%) | Monomer = 97.6 HMWS = 2.4 | Monomer = 99.2 HMWS = 0.8 |
| Replication competent AAV (in 1E+11 vg) | <10 rcAAV | <10 rcAAV |
| Residual Host Cell Protein (ng/mL) | 8.1 | <2.0 |
| Residual Host Cell DNA (ng DNA/E+13 vg) | < 0.1 | 2.5 |
| Residual baculovirus DNA/plasmid (copies/E+13 vg) | 2.0E+6 | 2.0E+11 |



SEC = size exclusion chromatography; HMWS = high molecular weight species

vg = vector genome

AAV Products from Both Processes Show Similar Activity Using a Functional (Enzymatic) Potency Assay



| Process | Relative Potency (%) |
|------------------------------|----------------------|
| Sf9-AAV | 100 |
| HEK-AAV | 87 |
| Assay variability is +/- 25% | |

Conclusions

- We thoroughly characterized and compared the final products (containing the same GMI) generated using an Sf9 and a HEK process in order to address the question of which is a better process
- Using developed processes, both methods yielded high quality vector with low amounts of impurities, a high % of full capsids, and low levels of post translational modifications
- Considerations/Caveats
 - Design of RepCap construct plays a significant role in high quality product from Sf9 system, and we have used an optimized design in this study.
 - Downstream process has some differences in buffer pH for the anion-exchange chromatography step.
 - Does not include long-read sequencing data
 - No in-vivo studies performed
- While there were minor differences in the product quality, the biological function was comparable for Sf9 and HEK derived products
- Sf9 had consistently higher yields and is our platform of choice, while we use HEK for indications requiring less drug product

Acknowledgements

Process Development

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